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**Plant and Soil**

An International Journal on Plant-Soil Relationships

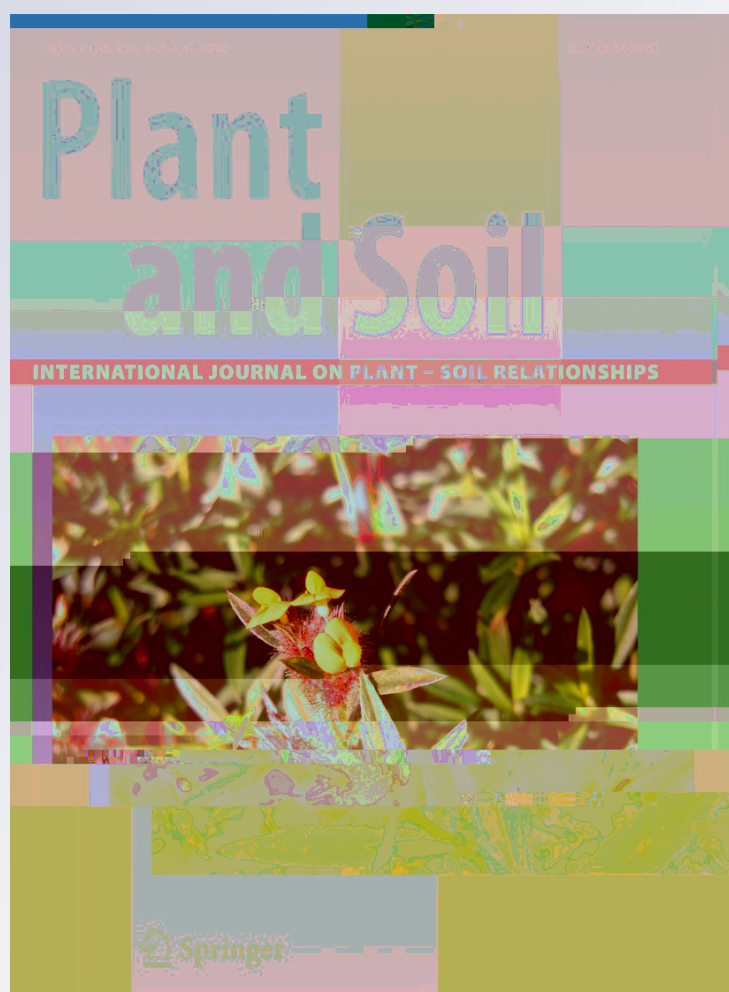
ISSN 0032-079X

Volume 425

Combined 1-2

Plant Soil (2018) 425:231-240

DOI 10.1007/s11104-018-3578-9



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# Intraspecific aggregation and soil heterogeneity: competitive interactions of two clonal plants with contrasting spatial architecture

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Received: 9 October 2017 / Accepted: 4 January 2018 / Published online: 7 February 2018  
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## Abstract

**Background and aims** Intraspecific aggregation of plant individuals can promote species coexistence by delaying competitive exclusions. However, such impacts may differ among species with contrasting spatial architecture and rely on the spatial distribution of resources.

**Methods** We grew a phalanx clonal plant *Carex neurocarpa* (with aggregated ramets) and a guerilla one *Bolboschoenus planiculmis* (with diffused ramets) in monocultures or in 1:1 mixtures with an even or a clustered distribution pattern of the two species in homogeneous or heterogeneous soils.

**Results** After 16 months, shoot biomass and ramet number were greater in mixtures than in monocultures in *C. neurocarpa*, but smaller in *B. planiculmis*. However,

the growth of neither *C. neurocarpa* nor *B. planiculmis* differed between even and clustered mixtures. Soil nutrient heterogeneity did not significantly affect the growth of either species, but increased relative yield of *B. planiculmis* and decreased that of *C. neurocarpa*.

**Conclusions** The relative importance of intra- vs. interspecific competition depends on the spatial architecture of plants, and soil nutrient heterogeneity slows down competitive exclusion by decreasing differences in competitive ability between plants. However, our results do not support the idea that intraspecific aggregation of individuals alters competitive interactions between species.

**Keywords** Aggregation · Clonal growth form · Clonal population · Competition · Environmental heterogeneity · Guerilla and phalanx

## Introduction

Intraspecific aggregation of plant individuals is a com-

processes. For instance, it may change light interception and water use (Mokany et al. 2008), soil nutrient accumulation (Derner and Briske 2001) and litter decomposition (Yu et al. 2011). Hence, spatial aggregation of conspecific individuals can alter the relative importance of intra- vs. interspecific competition, affecting species coexistence (Bolker et al. 2003; Houseman 2014; Lenssen et al. 2005; Stoll and Prati 2001; Tilman and Pacala 1993; Wassmuth et al. 2009). So far, studies testing the impact of spatial intraspecific aggregation of plant individuals have been mostly conducted in homogeneous environments (e.g. Lenssen et al. 2005; Monzeglio and Stoll 2008; Stoll and Prati 2001), without considering the inherent nature of environmental heterogeneity.

Natural environments are ubiquitously heterogeneous and essential resources for plant growth are commonly patchily distributed (Stuefer et al. 1996). A clonal plant may place more ramets in patches of higher resources by shortening inter-ramet distance (foraging strategy of a single clonal fragment, e.g. Dong et al. 2015; Peng et al. 2013; Slade and Hutchings 1987). A clonal plant may also increase the size of ramets in the higher resource patches by producing more leaves and roots, but show no change of inter-ramet distance (consolidate strategy of a single clonal fragment, e.g. Alpert and Mooney 1996; Birch and Hutchings 1994; de Kroon and Schieving 1990; Lovett-Doust 1987). When several independent clonal fragments grow together, they may all sense and thus put more new ramets and/or increase ramet size in higher resource patches (foraging or consolidate strategies of several independent clonal fragments, e.g. Day et al. 2003; Fransen et al. 2001; Liu et al. 2017; Wang et al. 2016). Such responses, in turn, increase intraspecific aggregation (Lara-Romero et al. 2016; Maestre et al. 2003; Maestre and Cortina 2002; Seabloom et al. 2005), and may further alter competitive interactions between intra- and interspecific individuals (Lara-Romero et al. 2016; Maestre and Reynolds 2007; Monzeglio and Stoll 2008; Skaer Thomason and Rice 2017; Wijesinghe and Hutchings 1999). We therefore hypothesized that environmental heterogeneity will enhance the impact of intraspecific aggregation on competitive interactions between plant species. While many studies have tested the impacts of either environmental heterogeneity or spatial aggregation of intraspecific individuals on plant growth and competitive interactions, few have considered these two impacts simultaneously.

Plant species vary greatly in their spatial architectures and two contrasting spatial architectures have been identified for clonal plants, i.e., phalanx and guerilla (Lovett-Doust 1981). Clonal plants with a phalanx architecture produce no or short spacers connecting adjacent asexual individuals (ramets), so that ramets of the same genetic individual (genet) are spatially highly aggregated (Humphrey and Pyke 1998; Navas and Garnier 1990; Ye et al. 2006). By contrast, clonal plants with a guerilla architecture produce long spacers so that ramets of the same genet are widely spaced (Humphrey and Pyke 1998; Navas and Garnier 1990; Ye et al. 2006). Phalanx plants are expected to show advantages in acquiring local resources and thus may have competitive advantages in more crowded (with a higher spatial aggregation of individuals), homogeneous environments (Humphrey and Pyke 1998; Navas and Garnier 1990; Lopp and Sammul 2017; Saiz et al. 2016; Ye et al. 2006). By contrast, guerilla plants may have an advantage in exploiting open or high resource patches in heterogeneous environments through foraging (i.e. selective placement of ramets in high resource patches), but may benefit less in closed, homogeneous environments (Humphrey and Pyke 1998; Navas and Garnier 1990; Saiz et al. 2016; Sammul 2011; Ye et al. 2006). We are not aware of any studies that have tested simultaneously effects of environmental heterogeneity and spatial aggregation on the growth and competitive interactions of plants with contrasting spatial architectures. We hypothesized that impacts of environmental heterogeneity and intraspecific aggregation are different in guerilla and phalanx plants so that they alter competitive interactions between phalanx and guerilla plants.

To test our hypothesis, we grew a phalanx plant *Carex neurocarpa* and a guerilla plant *Bolboschoenus planiculmis* in monocultures and mixtures. In mixtures, the plants were grown either with an even or a clustered distribution pattern of the two species, and in either homogeneous soils or in heterogeneous soils consisting of high and low nutrient patches. Specifically, we addressed the following questions. (1) Does intraspecific aggregation of individuals affect the growth and competitive interactions of the two plants? (2) Does soil nutrient heterogeneity affect the growth and competitive interactions of the two plants with contrasting spatial architecture? (3) Is there an interactive effect of soil nutrient heterogeneity and intraspecific aggregation on the growth and competitive interactions of the two plants? p23

## Materials and methods

### Plant species

Both the phalanx clonal plant *Carex neurocarpa* Maxim. and the guerilla clonal plant *Bolboschoenus planiculmis* (F. Schmidt) T. V. Egorova (synonym: *Scirpus planiculmis* F. Schmidt) are perennial sedges of the Cyperaceae family (Chen et al. 1999). *Carex neurocarpa* is a tussock-forming clonal plant and produces very short rhizomes (inter-ramet distance <1 cm) and ramets of the same clone are closely spaced (Chen et al. 1999). In contrast, *B. planiculmis* forms long rhizomes (inter-ramet distance is up to 17 cm) so that ramets of the same clone are widely spaced (Chen et al. 1999). Rhizomes of *B. planiculmis* can branch intensively (Xue et al. 2013). Ramet height of *C. neurocarpa* is 0.2 to 1.0 m and that of *B. planiculmis* is 0.6 to 1.0 m. These two species are widely distributed and often coexist in wetlands in China (Chen et al. 1999).

### Sampling and cultivation

On 15 June 2012, we collected more than 1800 ramets of *C. neurocarpa* and 1800 ramets of *B. planiculmis* from 20 natural communities along the north bank of Miyun reservoir in Beijing (40.533° N, 117.016° E). We then cut each ramet at 10 cm above shoot base and planted it in a small pot (10 cm in diameter) in an experimental garden (40.547° N, 117.010° E) a few kilometers away from the sampling places. After one month of cultivation, most of the ramets survived and produced new leaves. We then selected 864 similar-sized ramets of both *C. neurocarpa* and *B. planiculmis* and used them in the experiment described below. Initial biomass of the ramets was  $0.132 \pm 0.019$  g (mean  $\pm$  SE,  $n = 21$ ) for *C. neurocarpa* and  $0.119 \pm 0.014$  g (mean  $\pm$  SE,  $n = 31$ ) for *B. planiculmis*.

### Experimental design

We pressed 48 wooden frames (50 cm wide  $\times$  50 cm long  $\times$  30 cm deep) into the soil to a depth of 25 cm in the experimental garden. The distance between adjacent frames was at least 0.5 m. The soil inside the wooden frames was removed and replaced with the experimental soil described below. Each frame was thereafter referred to as a plot.

The experiment consisted of two levels of soil heterogeneity (homogeneous vs. heterogeneous) crossed with

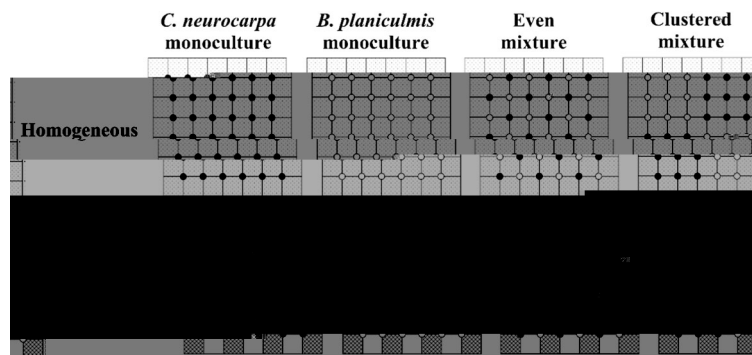
four levels of planting types (monoculture of *C. neurocarpa*, monoculture of *B. planiculmis*, an even mixture of *C. neurocarpa* and *B. planiculmis*, and a clustered mixture of *C. neurocarpa* and *B. planiculmis*; Fig. 1). There were eight treatments in total and six replicates (plots) in each treatment.

In the heterogeneous treatments, each plot was divided into 49 equal patches (7.1 cm  $\times$  7.1 cm each) using a metal grid. Patches were filled with either an 1:1 (v:v) mixture of potting compost (total N: 13.39 g kg<sup>-1</sup>; total P: 6.34 g kg<sup>-1</sup>; total K: 24.45 g kg<sup>-1</sup>) and sand (total N: 0.23 g kg<sup>-1</sup>; total P: 1.01 g kg<sup>-1</sup>; total K: 22.34 g kg<sup>-1</sup>) (hereafter refer to as high nutrient soil) or an 1:9 (v:v) mixture of the compost and sand (hereafter refer to as low nutrient soil; Fig. 1). High and low nutrient soil patches were filled alternately. In total, 25 patches were filled with high nutrient soil (high nutrient patches) and 24 patches with the low nutrient soil (low nutrient patches) in each plot in the heterogeneous treatments. Thus, the high and low nutrient soils differed greatly in total N and total P. In the homogeneous treatments, the plot was also divided into 49 equal patches (7.1 cm  $\times$  7.1 cm), and in each patch, we filled a 25:24 (v:v) mixture of the high and low nutrient soils (Fig. 1). In this way, the total amount of nutrients per plot was the same in the homogeneous and the heterogeneous treatments. After filling the plots, we removed the metal grid so that roots could grow across patches. Before filling the plots with the soil mixtures, we placed, at the bottom of each plot, a piece of non-woven fiber (50 cm  $\times$  50 cm) which is widely used as rooting cloth to block roots from growing outside the plot but allow vertical movement of water.

We then planted ramets of *C. neurocarpa* and *B. planiculmis* at the cross-points of the patches within each plot (Fig. 1). In monocultures we planted 36 ramets of *C. neurocarpa* or *B. planiculmis* within a plot, and in mixtures we planted 18 ramets of both *C. neurocarpa* and *B. planiculmis* (Fig. 1). In even mixtures, ramets of the two species were planted in alternate positions (Fig. 1). In clustered mixtures, the 36 planting positions in a plot were divided into four clusters with nine planting positions each, and nine ramets of each species were planted within a cluster (Fig. 1). Thus, in even and cluster mixtures the 18 ramets of both *C. neurocarpa* and *B. planiculmis* were conspecifically segregated and aggregated, respectively (Fig. 1).

The experiment was maintained for 16 months (from 17 July 2012 to 4 November 2013). During





**Fig. 1** Schematic representation of the experimental design. The experiment consisted of four homogeneous and four heterogeneous treatments with ramets grown in monocultures or mixtures, with the two species planted evenly or in clusters. In monocultures, 36 ramets of *Carex neurocarpa* (phalanx) or *Bolboschoenus planiculmis* (guerilla) were planted at the cross-points of the

patches within each frame, and in mixtures, 18 ramets of both species were intraspecifically segregated or aggregated within each frame. In the heterogeneous treatments, open and shaded patches represent patches with low and high nutrients, respectively. Black and open dots mark the positions where the ramets of *C. neurocarpa* and *B. planiculmis* were initially planted

the experiment, the mean precipitation from June to September was 329 mm in 2012 and 407 mm in 2013. Water was added to the plots when drought occurred in summer.

#### Harvest and measurement

Aboveground parts of each species were harvested at the end of experiment on 4 November 2013. We counted ramets of each species and harvested their aboveground shoots by cutting all plant material at ground level in each plot. For the guerilla plant (*B. planiculmis*) we counted ramets and harvested aboveground shoots in each type of soil patches (high vs. low nutrient patches) separately in the heterogeneous treatments. In the homogeneous treatments we also counted ramets of *B. planiculmis* and harvested the aboveground shoots in the same way as in the heterogeneous treatment, i.e. separately in patches that were located at the same places as the high and low nutrient soil patches in the heterogeneous treatment. As the ramets of the phalanx plant (*C. neurocarpa*) did not grow off the locations where it was planted, we harvested aboveground shoot biomass for this species in each plot but not separately for the two types of soil patches within each plot. Dry mass of all plant parts was determined after oven-dried at 70 °C for at least 48 h.

#### Data analysis

At the plot level, we first calculated shoot mass and ramet number per initial ramet of *C. neurocarpa* and

*B. planiculmis* separately in each plot. Since the growth of the two species in the mixtures was not independent, we performed separate two-way ANOVAs to test the effects of soil nutrient heterogeneity (homogeneous vs. heterogeneous) and planting type (monoculture vs. even mixture vs. clustered mixture) on the growth measures of each of the two species at the plot level. Following ANOVA, planned contrasts were conducted to further separate the effect of planting type into the effect of competition type [intra- vs. interspecific competition, i.e. monoculture vs. (even mixture plus clustered mixture)] and the effect of intraspecific aggregation of plant individuals (even mixture vs. clustered mixture).

To directly examine the competitive interaction between the two species, we calculated relative yield of each species by dividing its shoot mass per initial ramet in each mixture (even or clustered mixture) by mean shoot mass per initial ramet in monocultures across the six replicates. We used two-way ANOVA to test the effects of soil nutrient heterogeneity and intraspecific aggregation of plant individuals (even mixture vs. clustered mixture) on relative yield of each of the two species separately.

At the patch level, we first calculated shoot mass and ramet number per initial ramet per patch of the guerilla clonal plant *B. planiculmis* in both types of soil patches within the plots. We performed three-way ANOVA with repeated measures to test the effects of soil nutrient heterogeneity, planting type and patch type (high vs. low nutrient patches) within the plots on the growth of *B. planiculmis*. Following ANOVA, planned contrasts were conducted to further separate the effect

of planting type into the effect of competition type and that of intraspecific aggregation. In this analysis, patch type was treated as a repeated variable as data in the high and low nutrient soil patches in the same plot were not independent.

Before analysis, data of shoot mass and number of ramets at the plot level and data of shoot mass of *B. planiculmis* at the patch level were transformed to square root to improve normality and homoscedasticity. All analyses were performed with R (version 3.3.2; <http://www.r-project.org>) in RStudio (version 1.0.44; <http://rstudio.org>).

## Results

Shoot mass and ramet number were greater in mixtures than in monocultures in *C. neurocarpa* (Table 1a; Fig. 2a-b), but smaller in *B. planiculmis* (Table 1b;

Fig. 2c-d). However, intraspecific aggregation significantly affected the growth of neither *C. neurocarpa* nor *B. planiculmis* (Table 1; Fig. 2). Soil nutrient heterogeneity or its interactions with planting type did not significantly affect shoot mass or ramet number (Table 1; Fig. 2).

Relative yield was significantly greater in homogeneous than in heterogeneous soils in *C. neurocarpa* (Table 2a; Fig. 3a), but tended to be significantly smaller in *B. planiculmis* (Table 2b; Fig. 3b). Intraspecific aggregation of individuals or its interactions with soil nutrient heterogeneity did not significantly affect relative yield (Table 2).

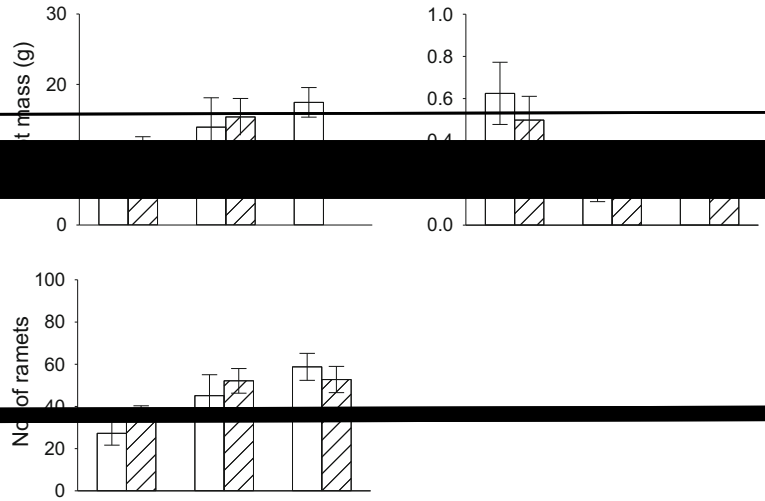
There were significant effects of soil nutrient heterogeneity  $\times$  patch type on the growth measures of *B. planiculmis* (Table 3). Shoot mass and ramet number of *B. planiculmis* were greater in the high than in the low nutrient patches in the heterogeneous soil treatments, but did not differ between the

**Table 1** Effects of soil nutrient heterogeneity (homogeneous vs. heterogeneous) and planting type (monoculture vs. even mixture vs. clustered mixture) on the growth of (a) *Carex neurocarpa* (phalanx) and (b) *Bolboschoenus planiculmis* (guerilla) at the plot level. The effect of planting type was further separated into the

effect of competition type [intra- vs. interspecific competition, i.e. monoculture vs. (even mixture plus clustered mixture)] and the effect of intraspecific aggregation (even mixture vs. clustered mixture) by planned contrasts

Effect	DF	Shoot mass <sup>1</sup>		No. of ramets <sup>1</sup>	
		<i>F</i>	<i>P</i>	<i>F</i>	<i>P</i>
(a) <i>Carex neurocarpa</i>					
Soil nutrient heterogeneity (H)	1	1.21	0.281	1.23	0.277
Planting type (P)	2	<b>4.22</b>	<b>0.024</b>	<b>6.65</b>	<b>0.004</b>
Competition type (C)	1	<b>7.51</b>	<b>0.010</b>	<b>12.04</b>	<b>0.002</b>
Intraspecific aggregation (A)	1	0.93	0.344	1.25	0.272
H $\times$ P	2	1.04	0.367	1.25	0.302
H $\times$ C	1	1.56	0.221	1.41	0.245
H $\times$ A	1	0.51	0.479	1.09	0.306
Residual	30				
(b) <i>Bolboschoenus planiculmis</i>					
Soil nutrient heterogeneity (H)	1	0.09	0.770	0.01	0.926
Planting type (P)	2	<b>5.65</b>	<b>0.008</b>	2.96	0.067
Competition type (C)	1	<b>9.57</b>	<b>0.004</b>	<b>4.57</b>	<b>0.041</b>
Intraspecific aggregation (A)	1	1.72	0.199	1.36	0.253
H $\times$ P	2	0.39	0.683	0.97	0.390
H $\times$ C	1	0.69	0.413	0.69	0.412
H $\times$ A	1	0.08	0.775	1.25	0.272
Residual	30				

<sup>1</sup>Data were transformed to square root. Values are in bold when  $P < 0.05$  and in italics when  $0.05 < P < 0.1$



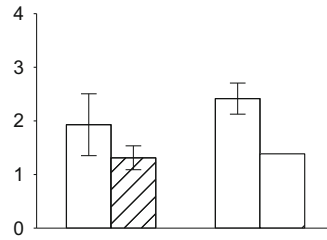
mirrored high and low nutrient patches in the homogeneous soil treatment (Table 3; Fig. 4).

## Discussion

Spatial architecture of plants can to some extent determine the uptake and the use of essential resources

(Ikegami et al. 2009; Lopp and Sammul 2017; Nacry et al. 2013; Sammul 2011; Xie et al. 2014; Ye et al. 2006) and thus may affect competitive interactions between plant species (Humphrey and Pyke 1998; Liao et al. 2014; Lopp and Sammul 2017; Sammul 2011; Schmid and Harper 1985). Clonal plants can differ greatly in horizontal spatial architecture based on the distribution pattern of ramets of the same clone (Lovett-Doust 1981; Ye et al. 2006). Phalanx clonal plants show an aggregated distribution of ramets and are supposed to show competitive advantages when directly competing with other species (such as guerilla clonal plants with diffused distribution of ramets; Humphrey and Pyke 1998; Lopp and Sammul 2017; Navas and Garnier 1990; Saiz et al. 2016). By contrast, guerilla clonal plants show advantages to explore open areas by means of foraging to increase resource uptake in heterogeneous environments (Cahill and McNickle 2011; Dong et al. 2015; Lopp and Sammul 2017; Rajaniemi and Reynolds 2004; Sammul 2011; Xue et al. 2013). Thus, the relative importance of intra- vs. interspecific competition is expected to differ between phalanx and guerilla clonal plants (Humphrey and Pyke 1998; Navas and Garnier 1990). We indeed found that the phalanx clonal plant *C. neurocarpa* and the guerilla clonal plant *B. planiculmis* showed contrasting responses to intra- vs. interspecific competition, i.e. the





growth of the *C. neurocarpa* was greater in mixtures than in monocultures, but that of *B. planiculmis* was the opposite. Our results thus provide support for the view that the spatial architecture of plants can affect the relative importance of intra- vs. interspecific competition and thus the competitive interactions between plant species (Humphrey and Pyke 1998; Navas and Garnier 1990; Saiz et al. 2016).

Individuals of many plant species are distributed in aggregation, and such intraspecific aggregation of individuals is expected to alter the competitive ability of plants (Hart and Marshall 2009; Lenssen et al. 2005; Monzeglio and Stoll 2005; Skaer Thomason and Rice 2017; Stoll and Prati 2001). However, our results did not show any evidence that spatial aggregation of conspecific individuals affected the growth and competition ability of

spatial aggregation of conspecific individuals due to the overwhelming suppression by the stronger competitors.

As expected, soil nutrient heterogeneity had little impact on the growth of the phalanx clonal

the two clonal plants, even though several previous studies showed that intraspecific aggregation benefited weaker competitors (Hart and Marshall 2009; Lamošová et al. 2010; Stoll and Prati 2001; Wassmuth et al. 2009). Intraspecific aggregation of plant individuals can influence plant growth because it can alter the relative importance of intra- vs. interspecific competition and slow down the competitive exclusion process. However, the phalanx clonal plant *C. neurocarpa* produced much more biomass than the guerilla clonal plant *B. planiculmis* at harvest. The overwhelming dominance of the phalanx clonal plant may have covered the potential positive effect of intraspecific aggregation on the competitive performance of the guerilla clonal plant. Consequently, we did not detect any impact of intraspecific aggregation. Therefore, the weaker competitor may not benefit from

individuals may alter their intra- and interspecific competitions in communities and thus affect their responses to environmental heterogeneity (Damgaard 2010; Lara-Romero et al. 2016; Monzeglio and Stoll 2008). Unexpectedly, however, we did not find an interactive effect of soil nutrient heterogeneity and spatial aggregation of conspecific individuals on the performance of the phalanx or the guerilla clonal plant. Our results suggest that the responses of clonal plants to soil nutrient heterogeneity may not depend on the spatial patterns of the individuals.

We conclude that the relative importance of intra- vs. interspecific competition depends on the spatial architecture of plants, and soil nutrient heterogeneity can slow down the competitive exclusion through decreasing the relative difference in competitive ability between plants. However, our results do not support the idea that intraspecific aggregation of plant individuals can alter competitive interactions between species.

**Acknowledgements** We thank Rui Zhu, Bi-Cheng Dong, Pu Wang, Yong-Yang Wang, Jia-Yuan Li, Yong-Hong Gao and Xing-Xing Jiang for assistance with the experiment, and Jasper van Ruijven and two anonymous reviewers for valuable comments on the earlier version of the manuscript. This work was supported by NSFC (31570413 and 31761123001).

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